

✓ **Is it bird flu ?**

- Type A specific TaqMan

✓ **Is it H5 or H7 ?**

- Subtype H5 and H7 specific TaqMan

✓ **Is it HP or LP strain ?**

- PCR & sequencing of HA cleavage region

✓ **( Is it something else ? What? )**

- TaqMan array (NDV, WNV, vvIBDV, other?)



# AI TaqMan technology transfer to state labs and evaluation of different instrument platforms

## Instruments

- Applied Biosystems ABI 7700 Sequence Detection System
- ABI 7500 Fast plate system
- Corbett Research Rotor-Gene Instrument
- Biorad iCycler
- Roche Lightcycler
- Cepheid Smartcycler

## Transfer of AI TaqMan assays to state labs

- Harmonized assay using SOP, primer & probe sequences and assay conditions provided by AAHL
- Positive and negative controls (inactivated virus in RLT) sent to state labs
- Labs to extract RNA from positive control and perform real-time PCR on serial diluted RNA to determine analytical sensitivity
- Coded (“blind”) samples of strong and weak positives and negatives sent to state labs for proficiency testing

## Influenza Virus HI titres

| Serum     | A/tern/Australia/75 H5N3 | A chicken/Vietnam/8/2004 H5N1 |
|-----------|--------------------------|-------------------------------|
| Anti-H5N3 | 640                      | 40                            |
| Anti-H5N1 | 640                      | 5120                          |

**This data indicates that using an antigen in the HI test that is not matched to the outbreak strain will result in reduced sensitivity of the test**



## Molecular diagnostics for AI (cont)

**A further diagnostic challenge is illustrated by the information in our slide of the evolution of the current H5N1, which shows considerable genetic reassortment.**

**Pandemic preparedness requires the capability to sequence the whole genome to detect reassortments that may lead to a virus with different properties for virulence or infectivity in poultry, humans or other animals**

AI viruses have genomes with 8 segments

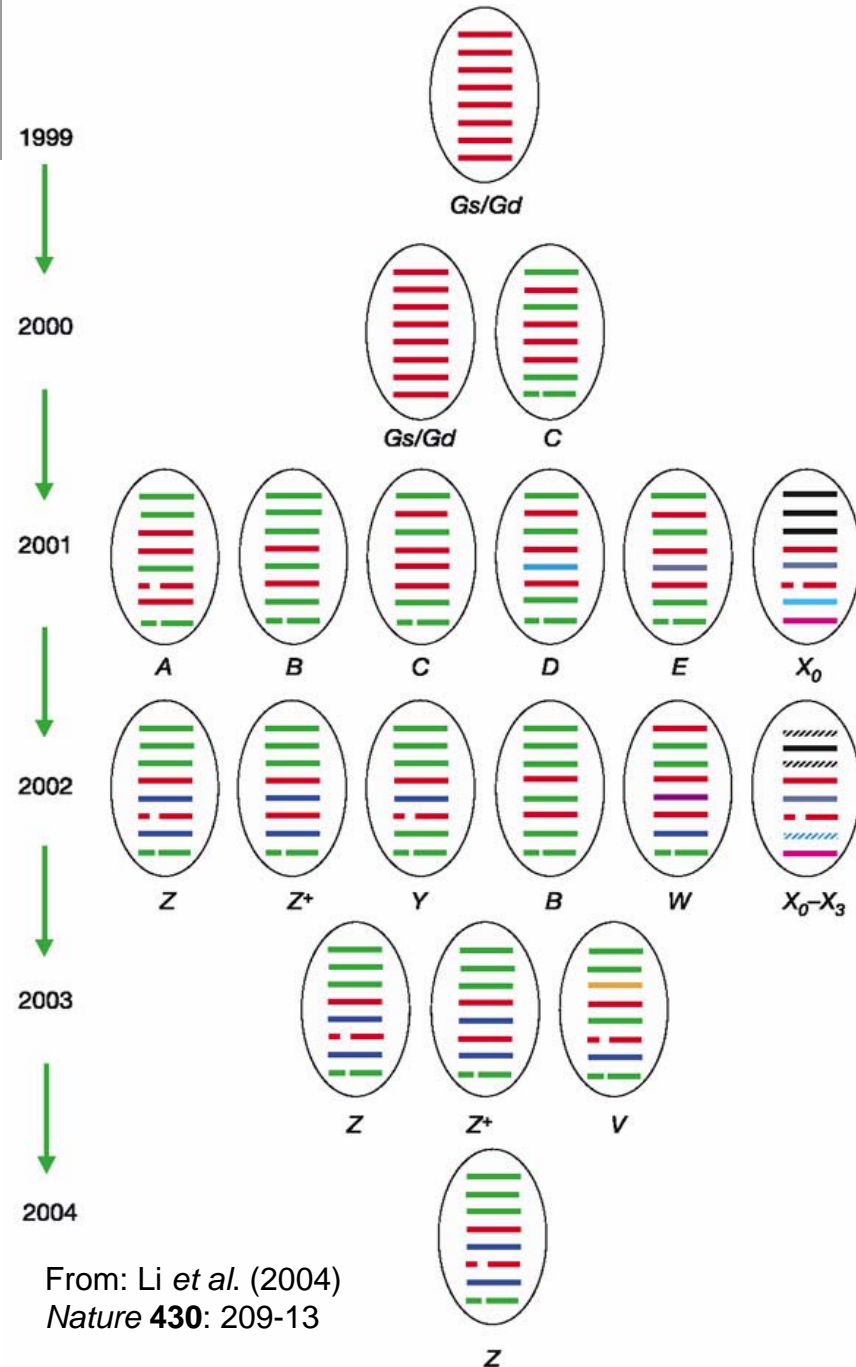
3 mechanisms for genetic evolution:

1. Point mutations and deletions
2. Intergenic recombination between segments
3. Genetic reassortment during dual infection of a cell

(Hence the evolution of H5N1 viruses as illustrated)

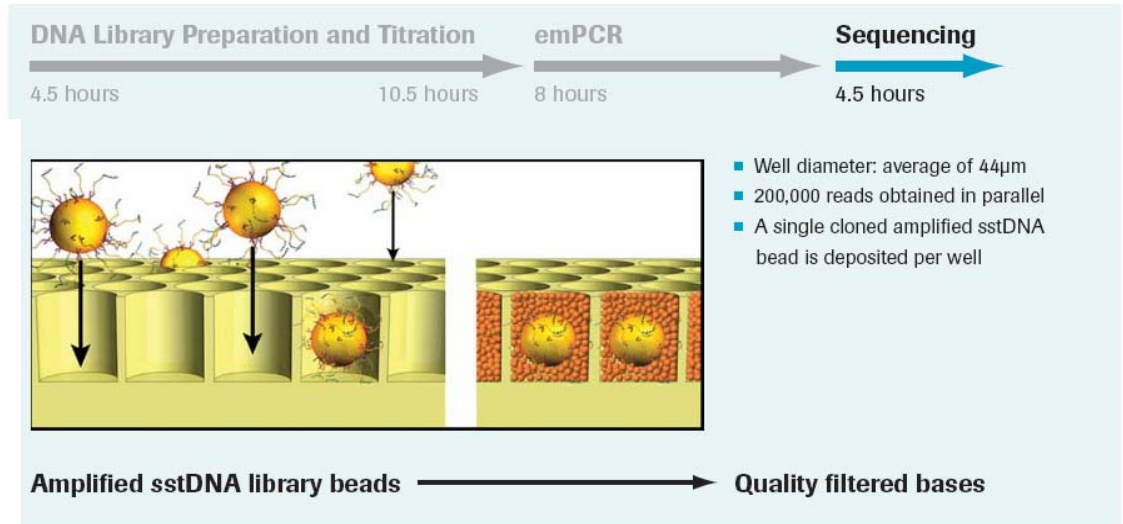
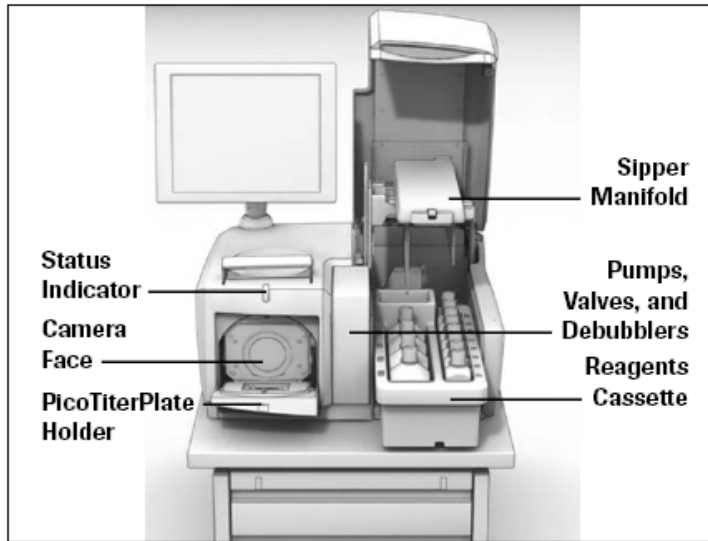
## Diagnostic challenges

- Primer/probe design
- Whole genome sequencing



From: Li *et al.* (2004)  
*Nature* **430**: 209-13

# Molecular Diagnostics: New technology for Rapid genome sequencing – not just a toy



**Single person in a day: 200,000 independent sequencing reactions → 2,000,000 bases**

**Article:** *Nature* advance online publication; published online 31 July 2005 | doi: 10.1038/nature03959  
**Genome sequencing in microfabricated high-density picolitre reactors**



# New technology will be adopted in reference laboratories

**Sequence more than 20  
million bases in 4-5 hrs**

**DNA library preparation-  
for RNA viruses need to  
add extra step of cDNA**

**Use to identify quasi-  
species**

**Potential applications to  
find new viruses**





## **Some examples of R&D responses to the Asian epidemic**



# Biosafety in the lab is of absolute importance



5 X 5 commercial ducks

6 weeks old

-ve by cELISA to AI antibody (all H types)

-ve to H5 by HI

$10^{-1}$  to  $10^{-5}$  infected AF

I/N; I/Oral; I/Ocular

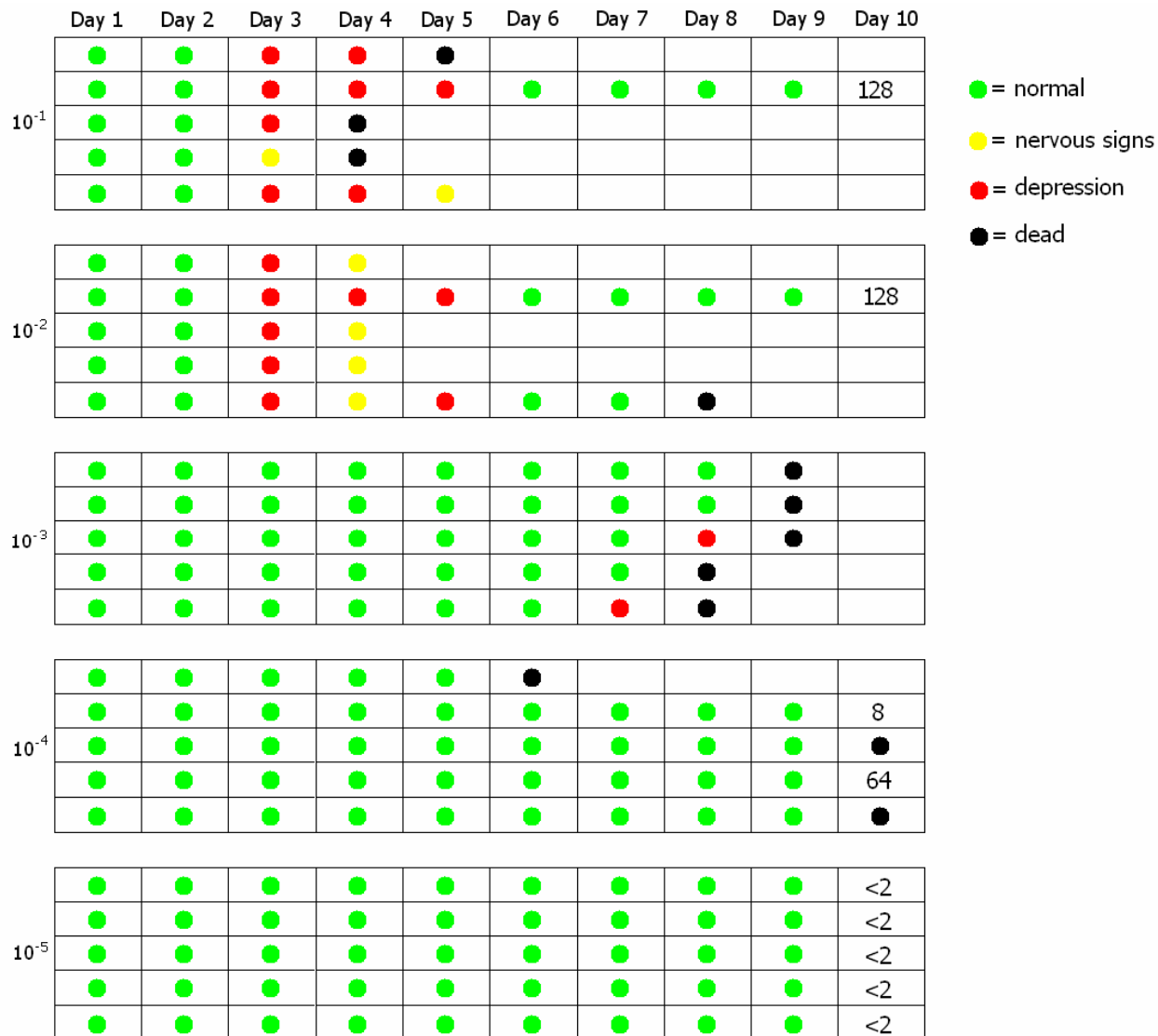
Observed for 10 days

- Clinical signs
- Virus excretion
  - Days 2, 3, 4, 7
- Antibody
  - Day 10

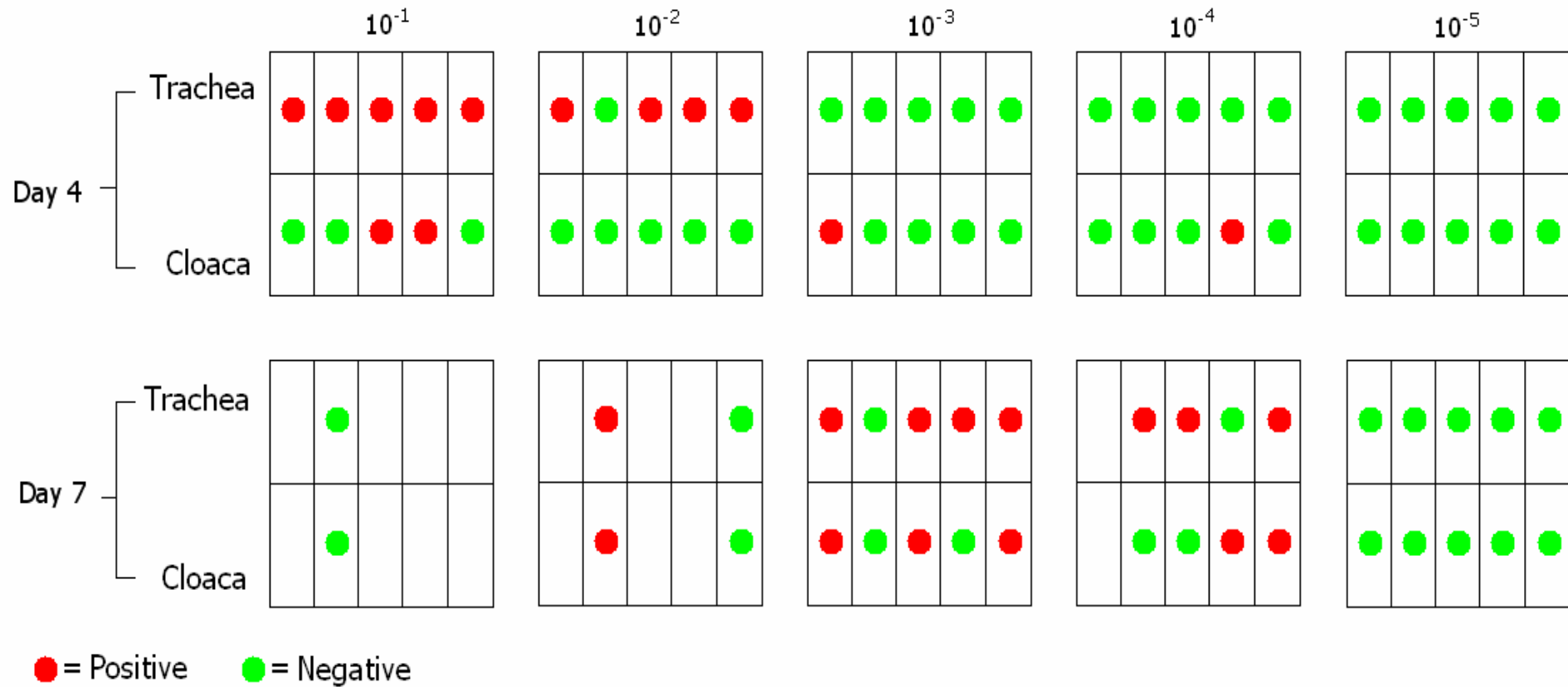




# Clinical observations – titration study



# Virus isolation – titration study



# Calculation of challenge dose

## An infected bird

- Exhibits clinical illness and/or
- Viral detection in tracheal or cloacal swabs on day 4 or day 7
- Titration end-point of  $10^{3.2}\text{EID}_{50}$

## Recommended $10^{-3}$ AF

- $10^{4.7}\text{EID}_{50}$  (equivalent to  $10^{1.5}\text{DID}_{50}$ )
- Over 30 duck infectious doses<sub>50</sub>



# Vaccine/H5N1 challenge study

## Control birds n=14

- PBS at day old and 3 weeks old

## Poulvac i-AI H5N9, H7N1 n=15

- 1<sup>o</sup> vaccination at day old and booster at 3wo

## Poulvac i-AI H5N3 n=15

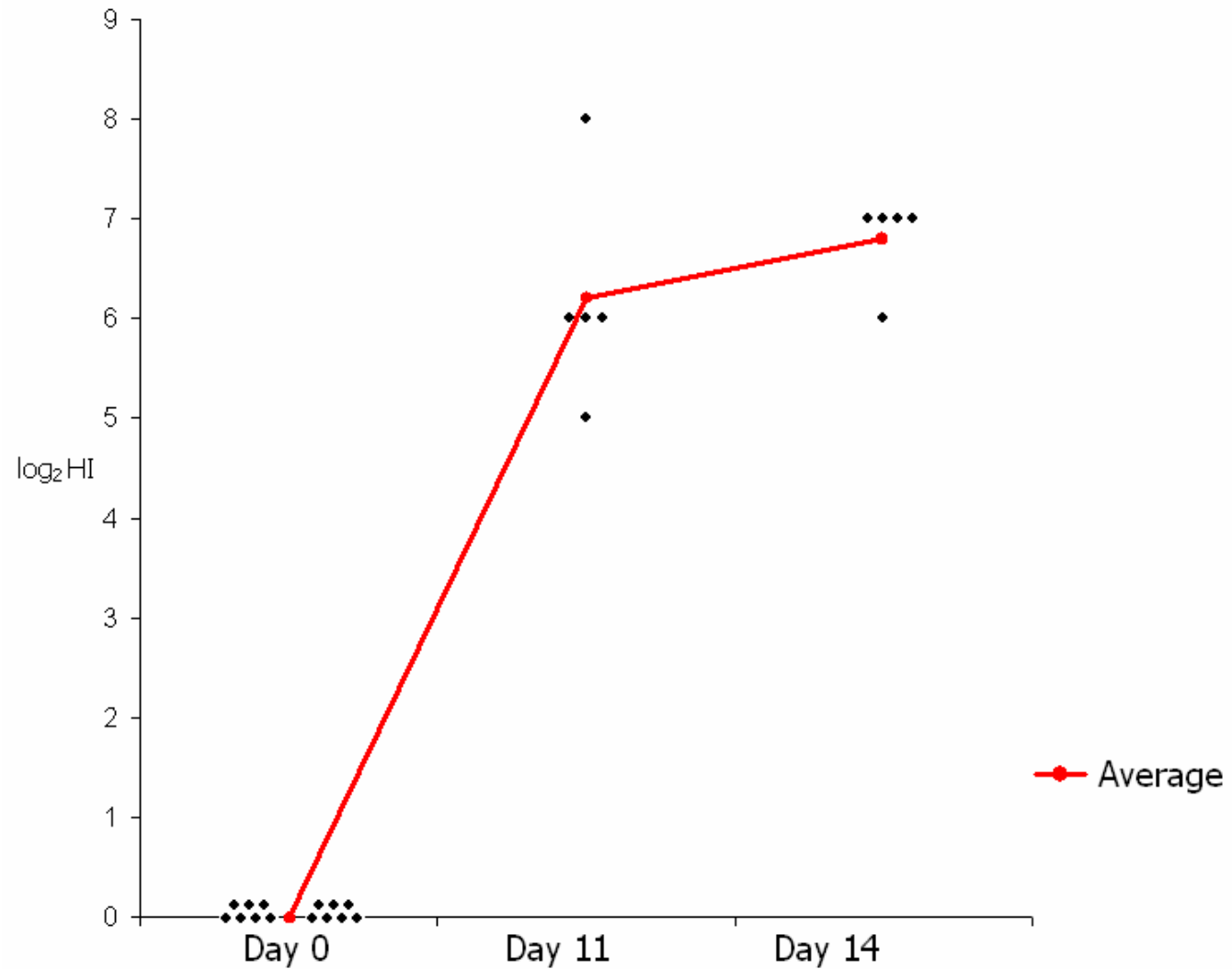
- 1<sup>o</sup> vaccination at day old and booster at 3wo

All birds challenged with H5N1 at 6wo

# Clinical observations – vaccine study



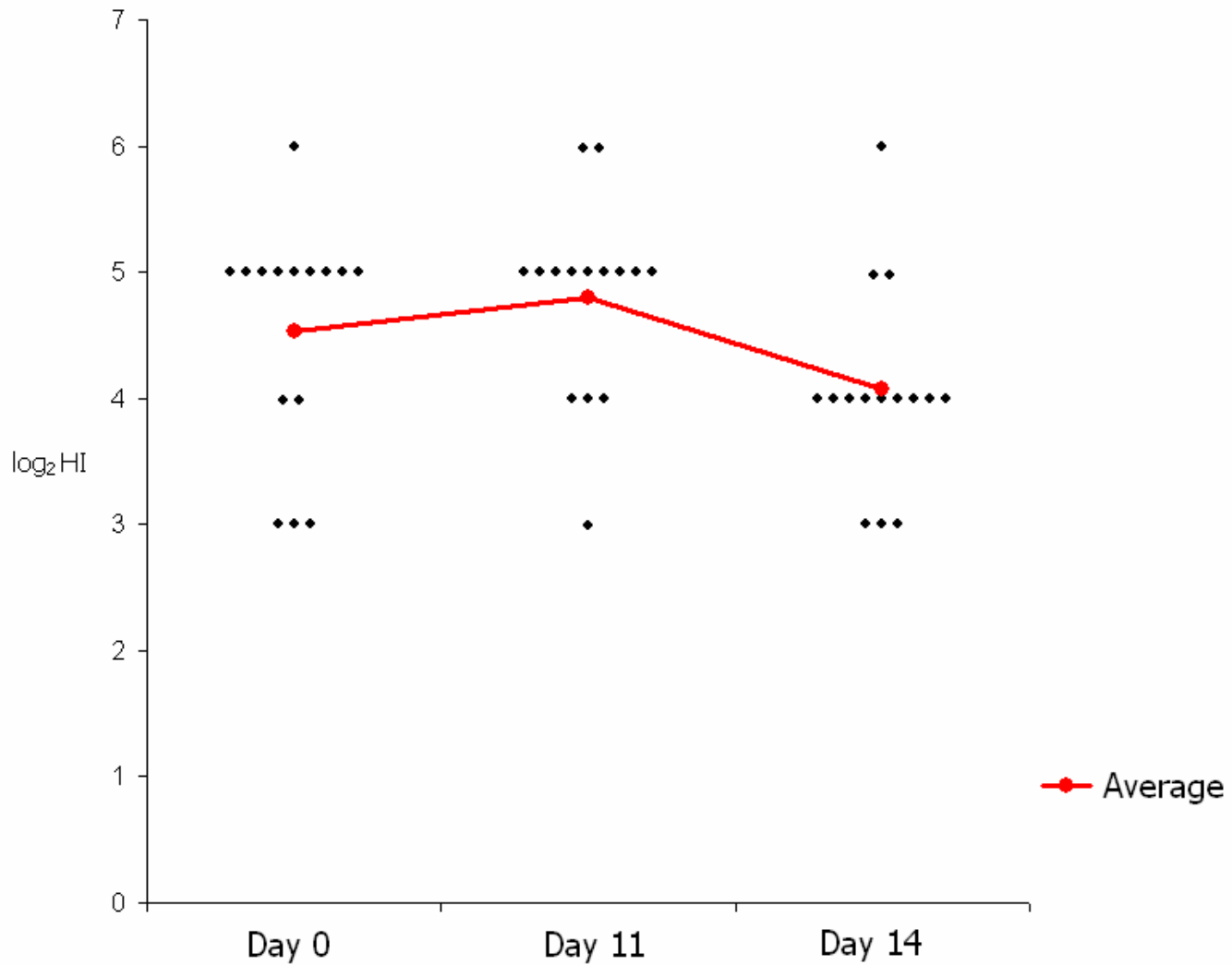
# Serology - controls





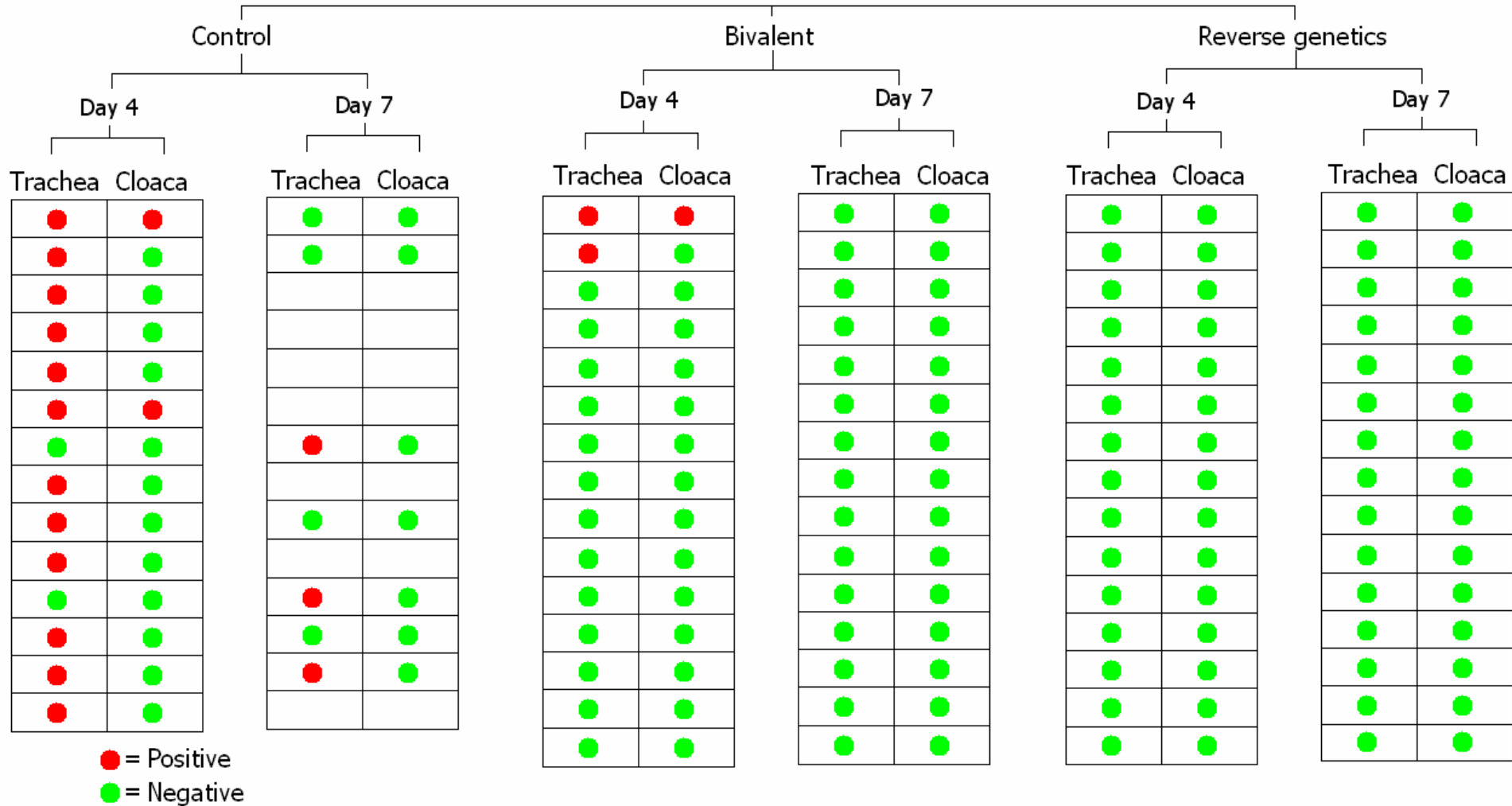


# Serology – H5N3 reverse genetics





# Virus isolation – vaccine study



## H5N9, H7N1 bivalent vaccine

- Protected ducks from H5N1 disease
- Serology data suggests infection occurred
- Shedding suppressed but not eliminated

## H5N3 reverse genetics vaccine

- Protected ducks from H5N1 disease
- Serology data suggests infection did not occur
- No viral shedding detected



# Point of Sampling Diagnostics

**Kits for rapid antigen/agent detection potentially play an important role**

**Which kit to use – will OIE registration help?**

**We need to learn how to use the kits, issues of**

- trained users (including sampling issues),
- Transport and storage issues
- validation of the diagnostic process
- Reporting issues – data capture and SOPs for response

**Test kits evaluated in this study were:-**

- Becton Dickenson Directigen Flu A & B,
- Biota Flu OIA,
- Anigen Rapid AIV Ag Test Kit,
- Anigen Rapid H5AIV Test Kit,
- Synbiotics Influenza Type A Antigen Test Kit



# Point of Sampling Diagnostics

## Analytical sensitivity

Dilutions of a stock virus (A/chicken/Vietnam/8/04 H5N1 with a titre of 108.1 EID<sub>50</sub>/ml) were made and 100 ul of each dilution added to the test kit sample buffer.

| Dilution | Directigen | Flu OIA | Anigen AIV | Anigen H5 | Synbiotics |
|----------|------------|---------|------------|-----------|------------|
| 1:5      | 4+         | 4+      | 4+         | Neg       | 4+         |
| 1:10     | 2+         | 2+      | 2+         | Neg       | 4+         |
| 1:50     | Neg        | Neg     | ?          | Neg       | 3+         |
| 1:100    | Neg        | Neg     | Neg        | Neg       | 1+         |
| 1:500    | Neg        | Neg     | Neg        | Neg       | Neg        |
| 1:1000   | Neg        | Neg     | Neg        | Neg       | Neg        |



# Point of Sampling Diagnostics

## **Tissues from H5N1 infected chickens**

Ten percent suspensions of eight tissues from experimentally infected birds were tested by the addition of 100 ul to the test kit sample buffer.

- All tissues were strongly positive in all kits.

## **Cloacal swabs from H5N1 infected chickens**

- The Directigen, Flu OIA and Anigen AIV kits all detected 3 of 4 swabs from experimentally infected chickens as strongly positive, with the 4th swab being weakly positive.
- The Anigen H5 failed to detect any of the swabs as positive.
- The Synbiotics kit has not been evaluated on swabs at this time.

All kits will be evaluated on tracheal swabs when clinical material is available.

## **CSIRO Livestock Industries**

### **Australian Animal Health Laboratory**

Peter Daniels

Assistant Director

Phone +61 3 5227 5014

Email [peter.daniels@csiro.au](mailto:peter.daniels@csiro.au)

Web <http://csiro.au/aah>



# Thank You

### Contact CSIRO

Phone 1300 363 400

+61 3 9545 2176

Email [enquiries@csiro.au](mailto:enquiries@csiro.au)

Web [www.csiro.au](http://www.csiro.au)